

## Chemiluminescence Immunoassay of Alpha-Fetoprotein

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### Abstract

Early diagnosis and treatment of cancer are the keys to improve patient survival rate. As the levels of tumor markers in serum are associated with the stages of tumors, reliable and sensitive determination of tumor markers plays a significant role in early cancer screening and evaluation. Protein detection methods, owing to the limited specificity of single markers in cancer diagnosis, multiplex immunoassays for analyzing a panel of tumor markers in complex serum samples to improve the diagnostic accuracy have attracted considerable attention.

A sensitive chemiluminescence (CL) imaging immunoassay method for detection of multiple tumor markers with high throughput, easy operation, and low cost was developed. The immunosensor was applied to detect  $\alpha$ -fetoprotein, carcinoma antigen 125, and carcinoembryonic antigen and to screen patients with liver cancer. The high throughput and acceptable stability, reproducibility, and accuracy showed good applicability of the proposed multiplex CL imaging immunoassay in clinical diagnosis.

**KEYWORDS:** chemiluminescence, immunosensors,  $\alpha$ -fetoprotein, cancer, liver.

### Introduction

The liver is a vital organ of vertebrates and some other animals. In the humans is located in the upper right quadrant of the abdomen, below the diaphragm. The liver has a wide range of functions, including detoxification of various metabolites, protein synthesis, and the production of biochemical necessary for digestion (1,3). There is currently no way to compensate for the absence of liver function in the long term, although liver dialysis techniques can be used in the short term (2,5,10).

The liver is a gland and plays a major role in metabolism with numerous functions in the human body, including regulation of glycogen storage, decomposition of red blood cells, plasma protein synthesis, hormone production, and detoxification. It is an accessory digestive gland and produces bile, an alkaline compound which aids in digestion via the emulsification of lipids. The liver's highly specialized tissue consisting of mostly hepatocytes regulates a wide variety of high-volume biochemical reactions, including the synthesis and breakdown of small and complex molecules, many of which are necessary for normal vital functions (4,13).

A tumor marker is a biomarker found in the blood, urine, or body tissues that can be elevated in cancer, among other tissue types (7,11,14). There are many different tumor markers, each indicative of a particular disease process, and they are used in oncology to

help detect the presence of cancer. An elevated level of a tumor marker can indicate cancer; however, there can also be other causes of the elevation.

Tumor markers can be produced directly by the tumor or by non-tumor cells as a response to the presence of a tumor. Most tumor markers are tumor antigens, but not all tumor antigens can be used as tumor markers (8,12).

Although mammography, ultra sono-graphy, computed tomography, magnetic resonance imaging scans, and tumor marker assays help in the staging and treatment of the cancer, they are usually not definitive diagnostic tests. The diagnosis is confirmed by biopsy.

Uses of tumor markers can broadly be classified as follows:

Diagnosis of specific tumor types, particularly in certain brain tumors and other instances where biopsy is not feasible.

As stated in the BMJ 2009, tumor markers should not generally be used for the purpose of diagnosis of cancers, as opposed to monitoring purposes in certain cancers, or in certain cases, screening purposes (9,13). The use of these tests without understanding their utility has resulted in inappropriate use of tumor marker blood tests, which has also resulted in further inappropriate over-investigation for cancers.

## MATERIALS AND METHODS

Materials and Reagents.

The capture antibodies (Ab1) and Ab2 of  $\alpha$ -fetoprotein (AFP) (mouse monoclonal antibodies, clones bsm-1021 and bsm-1022) were purchased from Beijing Biosynthesis Biotechnology Co. Ltd. (Beijing, China).

The corresponding electrochemiluminescent (ECL) immunoassay reagent kits for reference detection were provided by Roche Diagnostics GmbH (Germany).

Clinical serum samples were from the ambulatory and hospital person.

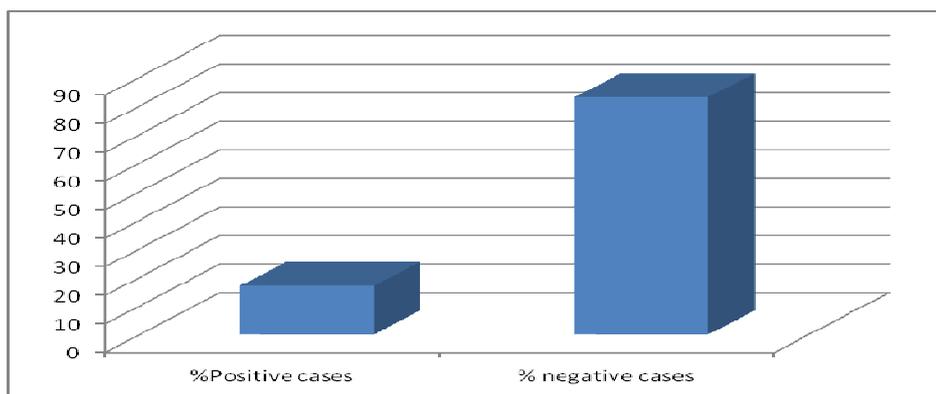
## RESULTS AND DISCUSSION

The collected CL intensity was proportional to the logarithm of analytic concentration over the ranges of  $(5.0 \times 10^{-5})$ –10 ng/mL or – 5,8 Unit/mL, for AFP. Positive detection rates in 102 serum samples by the CL imaging immunoassay are shown in Table 1.

The positive detection rates of AFP–CEA panel for 20 cases of liver cancer and CEA for 15 cases. As controls, no positive results were obtained in seven normal control samples. These results demonstrated the improved positive detection rates and diagnostic value for cancer screening, showing potential application in clinic diagnosis.

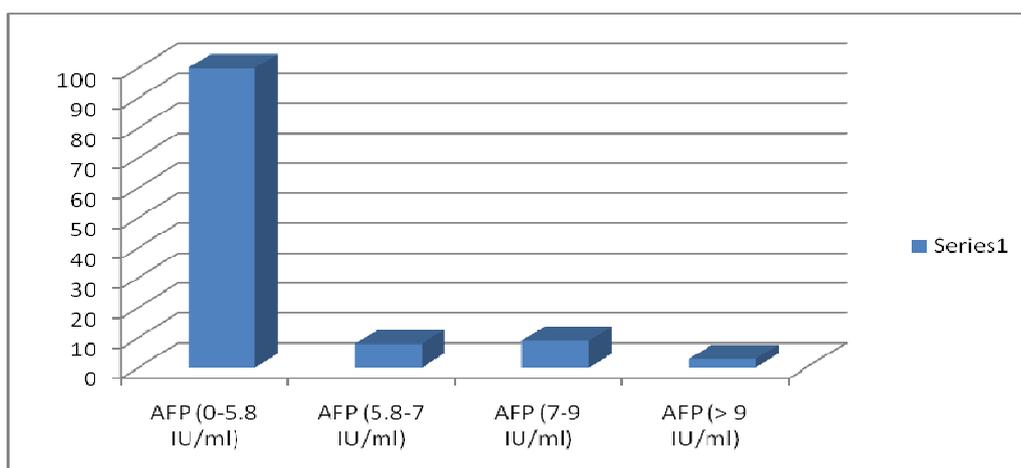
**Table 1.** Positive Detection of Clinical Sera

Cancer	Samples	Tumor marker	Positive cases
Liver cancer	120	AFP	20



**Figure 1:** Percentage of the cases with AFP

If we observed the figure 1, we can say that the most of the people who are tested for AFP resulted negative and only 16,6% of the total cases are positive by chemiluminiscence.



**Figure 2:** The cases with AFP

Basing on the AFP values we observed that the most of the positive cases are in values 7-9 IU/ml, and all the patients suffered from liver cancer.

## CONCLUSIONS

From our results we can say that a chemiluminescence immunoassay of multiple tumor markers is low cost, small consumption, simple operation, and high sensitivity for combination diagnosis of certain tumors. The availability of this system to measure simultaneously the levels of multimarkers in serum samples with sensitivity is a suitable to carry out large-scale screening of cancers in the early stage.

The level of relatively specific marker to certain tumor, for example, AFP to liver cancer, has the biggest difference from its cutoff value.

## REFERENCES

1. Ahmed, AS; Dew, T; Lawton, FG; Papadopoulos, AJ; Devaja, O; Raju, KS; Sherwood, RA (2007). "M2-PK as a novel marker in ovarian cancer. A prospective cohort study". *European journal of gynaecological oncology* **28** (2): 83–8.
2. Bagan P, Berna P, Assouad J, Hupertan V, Le Pimpec Barthes F, Riquet M (January 2008). "Value of cancer antigen 125 for diagnosis of pleural endometriosis in females with recurrent pneumothorax". *Eur. Respir. J.* **31** (1): 140–2.
3. Benesch, C; Schneider, C; Voelker, HU; Kapp, M; Caffier, H; Krockenberger, M; Dietl, J; Kammerer, U; Schmidt, M (2010). "The clinicopathological and prognostic relevance of pyruvate kinase M2 and pAkt expression in breast cancer". *Anticancer research* **30** (5): 1689–94.
4. Dennis Albert Casciato, Mary C. Territo Editors Dennis Albert Casciato, Mary C. Territo Contributor Mary C. Territo (2008): Manual of clinical oncology. Manual of Clinical Oncology Lippincott. Edition 6, 746.
5. Haug, U; Rothenbacher, D; Wentz, M N; Seiler, C M; Stegmaier, C; Brenner, H (2007). "Tumour M2-PK as a stool marker for colorectal cancer: Comparative analysis in a large sample of unselected older adults vs colorectal cancer patients". *British Journal of Cancer*.
6. Kaura, B; Bagga, R; Patel, FD (2004). "Evaluation of the Pyruvate Kinase isoenzyme tumor (Tu M2-PK) as a tumor marker for cervical carcinoma". *The journal of obstetrics and gynaecology research* **30** (3): 193–6.
7. Keshaviah, A; Dellapasqua, S; Rotmensz, N; Lindtner, J; Crivellari, D; Collins, J; Colleoni, M; Thurlimann, B et al. (2006). "CA15-3 and alkaline phosphatase as predictors for breast cancer recurrence: A combined analysis of seven International Breast Cancer Study Group trials". *Annals of Oncology* **18** (4): 701–8.
8. Kilpatrick, E. S; Lind, M. J (2009). "Appropriate requesting of serum tumor markers". *BMJ* **339**: 3111.
9. Koepke, John A. (1992). "Molecular marker test standardization". *Cancer* **69** (6 Suppl): 1578–81.
10. Kumar, Yogesh; Tapuria, Niteen; Kirmani, Naveed; Davidson, Brian R. (2007). "Tumour M2-pyruvate kinase: A gastrointestinal cancer marker". *European Journal of Gastroenterology & Hepatology* **19** (3): 265.
11. Leboeuf, R.; Langlois, Marie-France; Martin, Marc; Ahnadi, Charaf E.; Fink, Guy D. (2005). "'Hook Effect' in Calcitonin Immunoradiometric Assay in Patients with Metastatic Medullary Thyroid Carcinoma: Case Report and Review of the Literature". *Journal of Clinical Endocrinology & Metabolism* **91** (2): 361.
12. Osman N, O'Leary N, Mulcahy E, Barrett N, Wallis F, Hickey K, Gupta R (September 2008). "Correlation of serum CA125 with stage, grade and survival of patients with epithelial ovarian cancer at a single centre". *Ir Med J* **101** (8)
13. Schneider, J; Peltri, G; Bitterlich, N; Philipp, M; Velcovsky, HG; Morr, H; Katz, N; Eigenbrodt, E (2003). "Fuzzy logic-based tumor marker profiles improved sensitivity of

the detection of progression in small-cell lung cancer patients". *Clinical and experimental medicine* **2** (4): 185–91.

14. Wechsel, HW; Petri, E; Bichler, KH; Feil, G (1999). "Marker for renal cell carcinoma (RCC): The dimeric form of pyruvate kinase type M2 (Tu M2-PK)". *Anticancer research* **19** (4A): 2583–90.